

## Head of College Scholars List Scheme

## Summer Studentship

## **Report Form**

This report should be completed by the student with his/her project supervisor. It should summarise the work undertaken during the project and, once completed, should be sent by email to: <u>jill.morrison@glasgow.ac.uk</u> within four weeks of the end of the studentship.

1. Student

Surname: Rankin

Forename: Aime

E-mail address: 1002993r@student.gla.ac.uk

2. Supervisor:

Surname: Page

Forename: Roderic

E-mail address: Roderic.Page@glasgow.ac.uk

- 3. Research Project Report
  - 3.1 Project Title (maximum 20 words):

Are the parasites of marsupials ancient heirlooms or recent souvenirs?

3.2 Project Lay Summary (copied from application):

The parasites of an organism comprise a mixture of "heirlooms" and "souvenirs". Heirlooms are parasites retained from that species ancestor, and can be recognised by shared patterns of evolutionary history. Souvenirs are parasites that have been more recently acquired, for example as a result of a species moving into a new environment or evolving a new behaviour. This project seeks to determine what fraction of parasites on marsupial mammals are heirlooms reflecting their ancient past, and which parasites are souvenirs, perhaps of recent encounters with placental mammals.

3.3 Start Date: June 23rd

3.4 Original project aims and objectives (100 words max):

The null hypothesis is that the parasites of any marsupial mammal are more closely related to parasites found on other marsupials than to those on placental mammals. That is, marsupial parasites are heirlooms.

If we find parasites that are consistent with this hypothesis we can test a second hypothesis, namely that the marsupials and their parasites have cospeciated, by comparing evolutionary trees for both host and parasite. If cospeciation has occurred the trees should be more similar than we would expect due to chance.

If there are parasites that are shared between marsupial and placentals, we will test the hypothesis that sharing of parasites is related to degree of historical geographic isolation.

3.5 Methodology: Summarise and include reference to training received in research

methods etc. (250 words max):

Lists of marsupial host-parasite associations were obtained by searching through the Bionames database (http:bionames.org) or by searching the literature. Using phylogenies on Bionames and from the literature, close relatives of these parasites were found as well as their hosts and country of residence. Appropriate graphs and diagrams were then created using programs such as Mintab to summarise this data. Then the sequence database GenBank was searched for DNA sequences from those parasites. For each parasite sequence in GenBank, similar sequences were retrieved using BLAST and used to build а phylogeny (this process is automated using http://iphylo.org/~rpage/phyloinformatics/blast). For each resultant phylogeny the hosts for the parasites were identified, from GenBank or the literature. Through interpreting the tree, it was determined whether marsupial parasites tended to cluster together on the tree, suggesting evolutionary association with their hosts, or scattered across the tree, suggesting episodes of host switching. This process required the employment of various data mining and analysis skills relevant to evolutionary biology.

3.6 Results: Summarise key findings (300 words max). Please include any relevant tables or

images as an appendix to this report:

It was found that the close relatives of marsupial parasites, parasitized on a broad range of chordate groups. The majority of these were found on placental animals or mammalian animals in general (Figure 1). Only 5.5% of the parasites' relatives lived exclusively on marsupials and these came only from Australian Macropods (Figure 1). A small proportion of the parasites was found on Cephalopods, Insects or were free-living (Figure 1).

The range of countries in which these close relatives resided was equally as broad. The majority of the parasites' relatives (34%) had a worldwide presence, followed by 16.1% residing in Australia (Figure 2). Very few resided in South America and fewer still in New Guinea (Figure 2). However, for New Guinean parasites, the majority of their close relatives also resided there (Figure 2). Half of Australian parasites had relatives in Australia (Figure 2). Very few South American parasites had relatives in Australia (Figure 2).

When comparing the taxonomic distance between host and parasite pairs, there was a slight positive correlation between the taxonomic distance in host and parasite pairs (Figure 3). However, the taxonomic distance between the host pairs was greater than that of the parasite pairs in general (Figure 3). Several outliers were found and identified. These included *Ophidascaris robertsi* and *Hepatozoon vivernus*.

Similarly, a positive correlation between taxonomic distance of parasite pairs and geographic distance between host pairs was observed (Figure 4). Again, interesting outliers were identified. They included *Toxoplasma* and *Heterodoxus octoseriatus*.

The majority of parasite trees did not show obvious clustering of marsupial hosts. A surprising find was the occurrence of distantly related taxa placed within parasite groups. An example of this came from the phylogeny for *Parastrongyloides* (Figure 5). Within this tree, frogs, bears and even maize were included in the parasite's phylogeny (Figure 5). However, some trees did show clusters of marsupial parasites. These included *Sarcocystis, H. octoseriatus, Brachylaima dasyuri, Macropodinium ennuensis* and *M. yalabense* (Figures 6 to 9 respectively).

#### 3.7 Discussion (500 words max):

It was observed that few relatives of marsupial parasites were exclusive to marsupial hosts and that a majority were found to parasitize a great number of vertebrate groups (Figure 1). An explanation could be the presence of sampling error. Parasite phylogenies may not represent the diversity of parasites actually present. Not every parasite studied will be sequenced, so not every potential relative can be included in the tree. This could result in two parasites appearing more closely related than they are. For example, *Aggregata* appears as the closest relative of *H. vivernus* yet this would suggest a close relationship between marsupials and its host Cephalopods, which is not likely since there are no marine marsupials or for that matter, terrestrial Cephalopods.

Furthermore, the geographic distribution of the parasites was found to be diverse (Figure 2). Only parasites of New Guinea had the majority of relatives in the same area. This may be because the island has been relatively isolated since the end of the last ice age (Gascoigne, 1998). Regards the number of parasites found worldwide, it may be that the parasites are not host specific such as *Strongyloides* and can infect a range of vertebrates, hence their global presence. Again, sampling error would affect these results, so geographically distant parasites may not be as closely related to the marsupial parasites as suggested. This may explain why so few parasites of South American marsupials appear to have relatives in the same region.

As might be expected, the more taxonomically distant a parasite was from its closest relative, the more taxonomically distant the hosts of these parasites would be (Figure 3). Similarly, increasing geographic distance between host pairs correlated with the taxonomic distance between parasite pairs (Figure 4). Similar results of this nature have been found in other studies looking at helminth communities (Poulin, 2003).

The abundance of phylogenies not showing obvious clustering of marsupial hosts could be due to the sampling error discussed earlier. The occurrence of distantly related taxa in the tree itself may be down to using sequences from conserved genes or it could be that very short sequences were used. Additionally, it could be the case that the sequence used may have at some point integrated itself into the host's genome making it appear that the parasite and host were closely related. Therefore, an improvement to BLAST would be the addition of confidence values to give an impression of the reliability of the tree.

For the trees that did exhibit marsupial hosts clustered around groups of parasites, this could be evidence of cospeciation and further research should be carried out to test this. Studies on the protozoan *Sarcocystis* (Figure 6) have produced results suggesting that Sarcocystids from Australia and South America are monophyletic and have co-evolved with their marsupial hosts (Merino et al., 2010). Another theory has been that Boopiidae lice, including Australian *H. octoseriatus* (Figure 7), share a common ancestor with South American lice of marsupials (Barker, 1994). However, characters have been discovered to suggest a sister-group of bird lice and Boopiids and that the transfer of bird lice to marsupials creating the Boopiidae family is relatively recent (Barker, 1994). Many associations have been suggested for the genus *Macropodinium* (Figure 9). Coevolution and host switching have been observed while some speciation events are thought to have occurred due to vicariation in the Pleistocene, when marsupial populations split (Cameron and O'Donoghue, 2004). Another theory is that the associations of these ciliates with their hosts are not by co-descent but by resource tracking the diets of their hosts (Cameron and O'Donoghue, 2004). For each of these examples however, definite associations cannot be assumed until robust parasite and host phylogenies are formed.

To conclude, only a small number of the marsupial parasites investigated showed evidence of cospeciation but of these parasites, most were present in literature which reinforced their status as heirlooms. For the remainder of the parasites investigated, it was hard to establish their provenance as there was much variety in the taxonomic and geographic distances between themselves and the

assumed close relatives. Perhaps most likely due to sampling error, further study would be benefitted by closing the gaps in our sequence databases.

Cameron, S. L., 2004. Morphometric and Cladistic Analyses of the Phylogeny of Macropodinium (Ciliophora: Litostomatea: Macropodiniidae). *Acta Protozoologica*, 43 (1), pp.43-53.

Gascoigne, I., 1998. *Papua New Guinea (Cultures of the World)*. 2<sup>nd</sup> ed. New York: Benchmark Books.

Merino, S., Martinez, J., Vasquez, R. A. and Slapeta, J., 2010. Monophyly of Marsupial Intraerythrocytic Apicomplexan Parasites From South America and Australia. *Parasitology*, 137, pp.37-43.

Poulin, R., 2003. The decay of similarity with geographical distance in parasite communities of vertebrate hosts. *Journal of Biogeography*, 30 (10), pp.1609-1615.

4. Reflection by the student on the experience and value of the studentship (300 words max):

As a Zoology student hoping to forge a career in academia, I found the studentship provided by the HoCSLS an invaluable introduction to the world of research in the field of taxonomy.

Although the process of designing and carrying out successful experiments is prominent in the current curriculum, it is limited to the subject areas being taught at the time as well as the resources and time available during term time. By applying for the studentship, I was able to pursue an area of zoology that interested me and conduct a study over a reasonable timescale. I believe that this is very good experience for carrying out the fourth year honours project and beyond.

Another advantage that the studentship has brought has been the opportunity to learn new skills from an established researcher in the field, such as building and interpreting phylogenies. Such skills are not focused on in the practical elements of my course so by learning them through the studentship it has greatly added to my learning experience at the university.

To conclude, the HoCSLS studentship has been a very enjoyable experience that has allowed me to expand upon my current studies and has given me confidence to pursue a career in research once I graduate.

5. Dissemination: (note any presentations/publications submitted/planned from the work):

A full report will be published on Figshare in the near future

(http://figshare.com/authors/Aime%20Rankin/434982).

6. Signatures: Supervisor

Date

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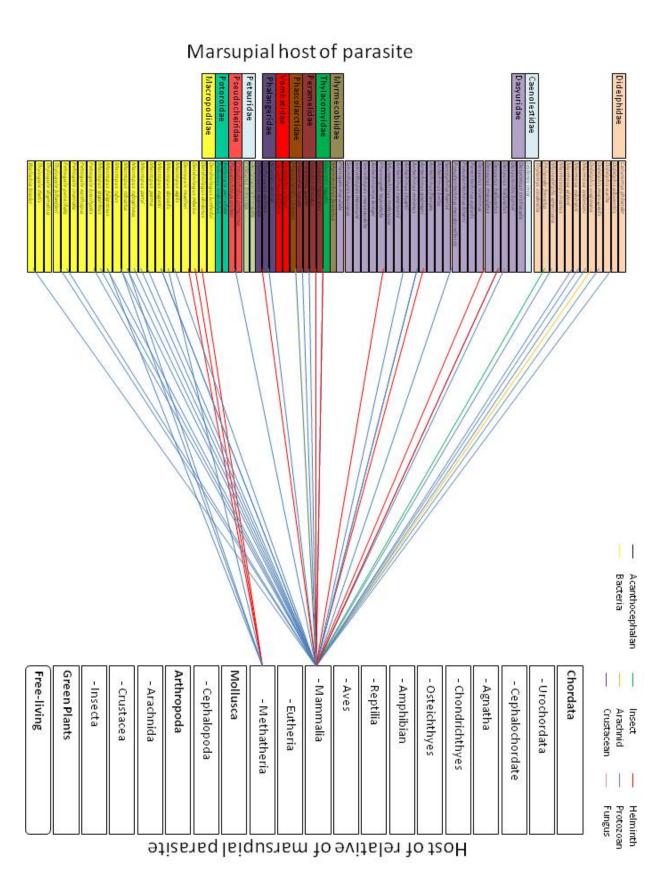
Date

Aime Rankin

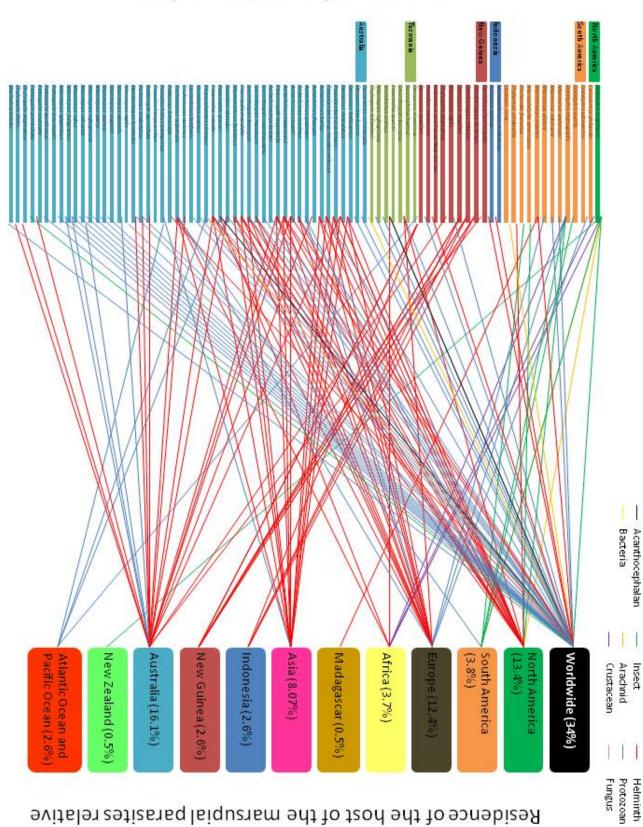
Student

22/07/2013

Figure 1: Diagram showing marsupial species and the hosts of the close relatives of their parasites. The colour of the line indicates the group to which the parasite belongs.

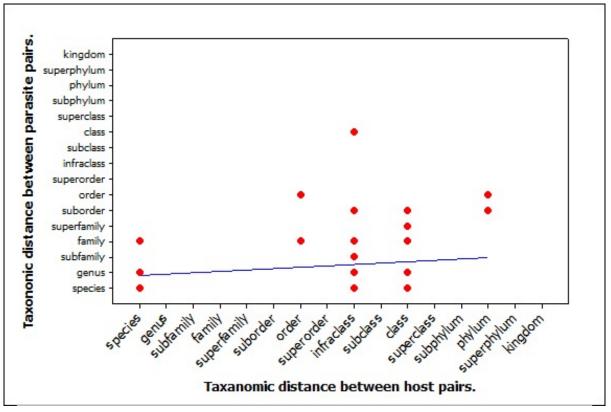


#### Appendix



Marsupial hosts country of residence

**Figure 2:** Diagram depicting the nationality of a marsupial and the country to which the host of their parasites closest relative lives. The colour of the line indicates the group to which the parasite belongs.



**Figure 3**: Graph depicting the taxonomic distance between marsupial parasites and their closest relatives, against the taxonomic distance between the marsupial host and the host of the parasites closest relative.

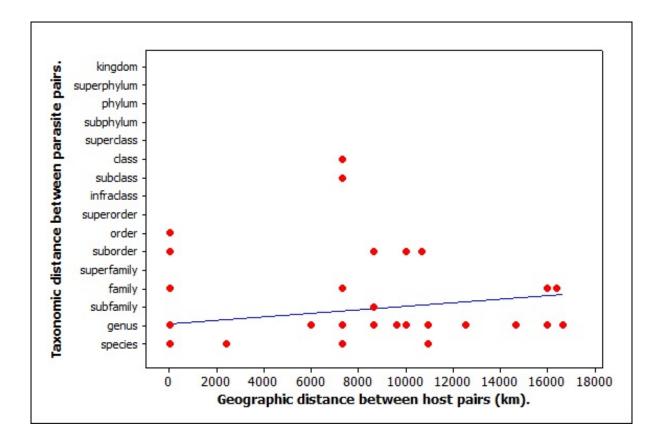
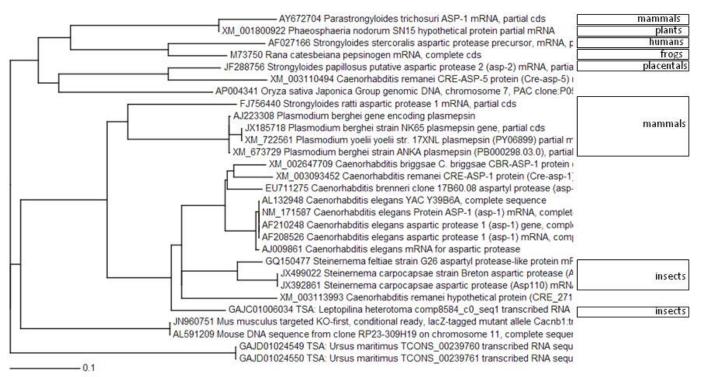


Figure 4: Graph depicting the taxonomic distance between marsupial parasites and their closest relatives, against the geographic distance between the marsupial host and the host of the parasites closest relative.

## Hosts of parasite



#### Figure 5: Phylogeny of Parastrongyloides generated by BLAST

## Hosts of parasite

| EU514791 Sarcocystis sp. cyst type III ex Anser albifrons 28S ribosomal RNA gene, partial sequence<br>EU514793 Sarcocystis sp. cyst type III ex Anser anser 28S ribosomal RNA gene, partial sequence   | vertebrate         |
|--|--------------------|
| EU553479 Sarcocystis sp. cyst type II ex Anas platyrhynchos isolate DA1 28S ribosomal RNA gene, partial sequen   | birds              |
| GU188426 Sarcocystis rileyi 28S ribosomal RNA gene, partial sequence<br>AF372656 Sarcocystis lindsayi large subunit ribosomal RNA gene, partial sequence<br>AF389341 Sarcocystis cf. falcatula large subunit ribosomal RNA gene, partial sequence<br>AF092927 Sarcocystis neurona 28S large subunit ribosomal RNA gene, complete sequence  | marsupials         |
| AF012883 Sarcocystis muris 28S large subunit ribosomal RNA gene, complete sequence<br>AF044251 Frenkelia glareoli large subunit ribosomal RNA gene, complete sequence<br>AF044252 Frenkelia microti large subunit ribosomal RNA gene, complete sequence<br>EU553480 Sarcocystis cornixi isolate V1 28S ribosomal RNA gene, partial sequence<br>FJ232949 Sarcocystis sp. ex Columba livia 28S ribosomal RNA gene, complete sequence<br>EU514792 Sarcocystis sp. cyst type I ex Anser albifrons 28S ribosomal RNA gene, partial sequence<br>U85705 Isospora felis 28S large subunit ribosomal RNA gene, partial sequence<br>DQ227420 Besnoitia besnoiti from Israel 18S ribosomal RNA gene, partial sequence; internal trai<br>DQ227419 Besnoitia besnoiti from Spain 18S ribosomal RNA gene, partial sequence; internal tra<br>AY833646 Besnoitia besnoiti from Portugal 18S ribosomal RNA gene, partial sequence; internal trai<br>AY616164 Besnoitia tarandi large subunit ribosomal RNA gene, partial sequence | placentals         |
| AF076900 Besnoitia besnoiti large subunit ribosomal RNA gene, complete sequence<br>AF489697 Besnoitia darlingi large subunit ribosomal RNA gene, partial sequence  | marsupials         |
| AF159240 Hammondia heydorni 28S large subunit ribosomal RNA gene, complete sequence<br>AF001946 Neospora caninum large subunit ribosomal RNA gene, complete sequence   | placentals         |
| AF076901 Toxoplasma gondii large subunit ribosomal RNA gene, complete sequence<br>L25635 Toxoplasma gondii strain RH 5.8S ribosomal RNA and ribosomal RNA large subunit genes<br>X75453 T.gondii (strain P) rDNA for 17s,5.8s,26s,and 5s ribosomal RNA<br>X75429 T.gondii (RH) 17S, 5.8S, 26S and 5S rRNA genes<br>AF101077 Hammondia hammondi 28S large subunit ribosomal RNA gene, complete sequence   | vertebrate         |
| AF237616 Sarcocystis zamani large subunit ribosomal RNA gene, parti  | Snakes and rodents |

## Hosts of parasite

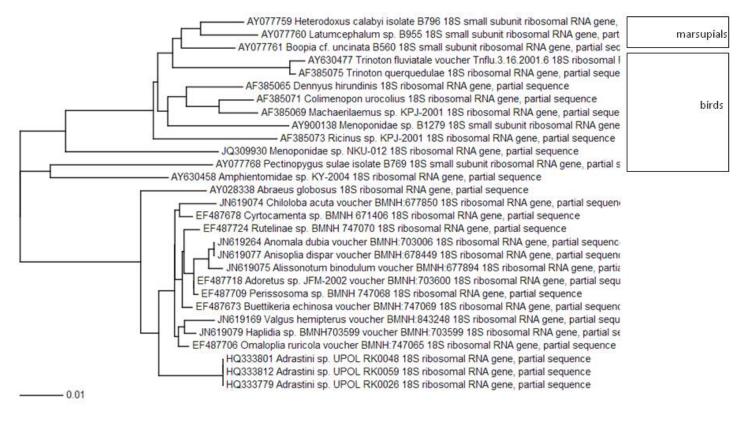


Figure 7: Phylogeny of H. octoseriatus generated by BLAST.

## Hosts of parasite

| marsupials and rodents  | AY222167 Brachylaima sp. Australia-PO-2003 28S ribosomal RNA gene, partia<br>DQ060330 Brachylaima virginianum 28S ribosomal RNA gene, partial s   |
|-------------------------|---|
| rodents                 | AF184262 Brachylaima thompsoni 28S ribosomal RNA gene, partial sequence<br>AB494468 Urogonimus macrostomus gene for 28S ribosomal RNA, partial sequence<br>JQ241173 Urogonimus macrostomus 28S ribosomal RNA gene, partial sequence<br>AY222168 Urogonimus macrostomus 28S ribosomal RNA gene, partial sequence<br>AY222169 Leucochloridium perturbatum 28S ribosomal RNA gene, partial sequence<br>AY222170 Zeylanurotrema spearei 28S ribosomal RNA gene, partial sequence<br>AB693170 Posthodiplostomum sp. NSMT:PI_5926 genes for 18S rRNA, ITS1, 5.8S rRNA<br>JF820607 Alaria taxideae isolate 3 28S ribosomal RNA gene, partial sequence<br>JF820605 Alaria taxideae isolate 1 28S ribosomal RNA gene, partial sequence |
| birds                   |   |
| amphibians              |   |
| birds and fish          |   |
| mustelids               |   |
| amphibians              | JF820597 Apharyngostrigea pipientis isolate 1 28S ribosomal RNA gene, partial sec   |
| fish                    | AY222172 Ichthyocotylurus erraticus 28S ribosomal RNA gene, partial sequence<br>HM114365 Hysteromorpha triloba 28S ribosomal RNA gene, partial sequence<br>AY222171 Cardiocephaloides longicollis 28S ribosomal RNA gene, partial sequence<br>AB551568 Liolope copulans gene for 28S ribosomal RNA, partial sequence<br>FJ609423 Clinostomum phalacrocoracis voucher 16/08-2TA 18S ribosomal RNA gene, inte   |
| birds                   |   |
| amphibians              |   |
| fish                    |   |
| birds                   | GQ339114 Clinostomum sp. 'cutaneum' voucher 129/09 18S ribosomal RNA gene, int<br>FJ609421 Clinostomum sp. 'cutaneum' voucher 118/07 18S ribosomal RNA gene, inte   |
|                         | AY222175 Clinostomum sp. Australia-PO-2003 28S ribosomal RNA gene, partial sec<br>AY858877 Clinostomid sp. W5005 28S ribosomal RNA gene, partial sequence   |
| fish                    | FJ609420 Clinostomum complanatum voucher 297/02 18S ribosomal RNA gene, inte<br>AY222176 Clinostomum sp. USA-PO-2003 28S ribosomal RNA gene, partial seguer   |
| reptiles                | FJ550134 Spirorchidae sp. CMA-2009 28S ribosomal RNA gene, partial sequen   |
| fish                    | FJ788481 Intusatrium robustum 28S large subunit ribosomal RNA gene  |
| reptiles and amphibians | JF820592 Telorchis bonnerensis isolate 3 18S ribosomal RNA gene pa  |

Figure 8: Phylogeny of Brachylaima dasyuri generated by BLAST.

# Host of parasite

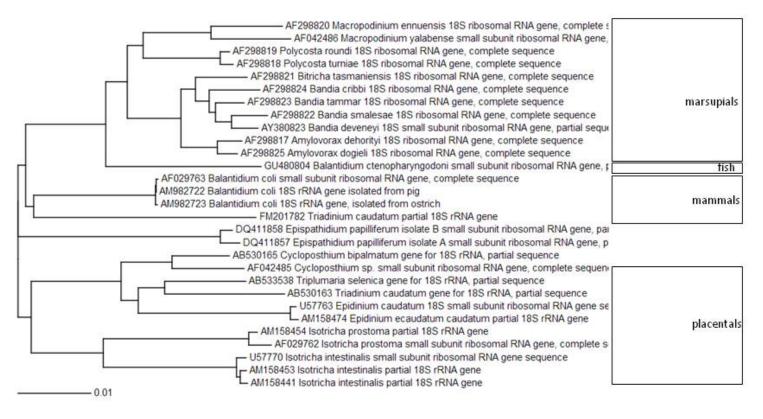


Figure 9: Phylogeny of Macropodinium ennuensis and M. yalabense generated by BLAST.